

In Vitro Evaluation of New, Potent, and Selective V₂ Receptor Agonists

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Introduction

The vasopressin analogue desmopressin (dDAVP, **1**) is a potent V₂ receptor agonist that also activates the related V_{1b} receptor [1]. dDAVP is approved in many countries for the treatment of diabetes insipidus, primary nocturnal enuresis, nocturia, and coagulation disorders including hemophilia A and von Willebrand's disease.

In search of novel, potent, selective and pharmacologically useful peptidic V₂R agonists, we synthesized a series of C-terminally truncated [Val⁴]dDAVP (**2**) [2] analogs modified in positions 2, 3, 7 and/or at the disulfide bridge. The peptides were evaluated for *in vitro* potency at the human V₂ receptor (hV₂R) and selectivity versus related receptors (hV_{1a}R, hV_{1b}R, hOTR). Here we present comprehensive *in vitro* data for the new compounds and describe synthetic methods used to prepare the analogues.

Results and Discussion

Analogues **3-24** (Table 1) were synthesized by a combination of solid and solution phase chemistry. The linear precursors of compounds **3-13**, **22** were assembled on H-Aaa-O-2-CITrt resins by standard Fmoc chemistry using DIC/HOBt mediated couplings. The carba thioether modifications of the disulfide bridge (X or Y = CH₂, Figure 1) were introduced to the peptide sequence by coupling Fmoc-

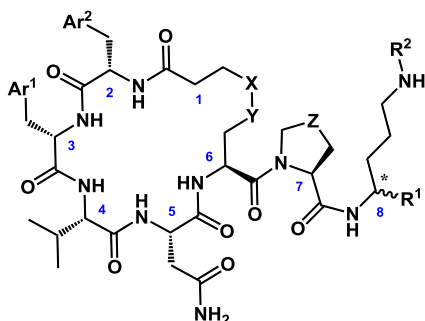


Fig. 1. General structure of new analogues **3-24**. Sequence positions numbered at α -carbons.

Cys((CH₂)₃-COOtBu)-OH or Fmoc-Hcy((CH₂)₂-COOtBu)-OH [3]. Fully protected peptide C-terminal acids were cleaved with 30% HFIP/DCM. For compounds **3-10** the carboxylic group was reduced to the hydroxymethyl group using mixed anhydride method [4]. For compounds **11-13** the C-terminal acids were coupled with agmatine and for analogue **22** the linear fragment was coupled with H-D-Arg(Pbf)-NEt₂. The protecting groups were removed with the TFA/TIS/H₂O 95/2.5/2.5 cocktail and the linear peptides were cyclized in DMF using HBTU/DIPEA method. The linear precursor of peptide **14** was assembled on 1,4-diaminobutane-2-CITrt resin. After cleavage from the resin the C-terminal amino function was temporarily protected with the TFA resistant Z(2-Cl) group. The peptide was deprotected and cyclized as described above. The Z(2-Cl) group was removed with TMSBr/thioanisole/TFA (1/1/6) [5]. The linear precursors of compounds **15-21**, **23** and **24** were synthesized on BAL resin which was reductively alkylated with an appropriate primary amine prior to the peptide assembly. The peptides were cleaved with concomitant side chain protecting group removal using the TFA cocktail and cyclized as described above. All peptides were purified by preparative HPLC and lyophilized.

The pharmacological profile of **1** was determined in *in vitro* assays and was consistent with the literature data [1]. **1** was particularly selective vs. the V_{1a}R (>1000-fold) presumably due to the desamino modification [6]. The Val⁴ analogue of **1** ([Val⁴]dDAVP, **2**) has been reported to be more potent and selective than **1** in rat *in vivo* models [7]. This profile is consistent with our *in vitro* studies at the human receptors (Table 1). Manning, et al. investigated the impact of the C-terminal Gly residue removal on the antidiuretic activity of **1**, **2** and related peptides in rats [7]. The corresponding desglycine analogues retained 10 - 50% initial antidiuretic activity, suggesting C-terminal truncation could be a good strategy to design novel V₂R agonists.

Table 1. Structure and in vitro pharmacological profile of analogs 1-24.

Analog	Structure ^a		Config of *C	hV ₂ R		Selectivity ^b vs. receptor		
	Ar ¹	R ¹		EC ₅₀ (nM)	E _{max} (%)	hV _{1a} R	hV _{1b} R	hOTR
1		dDAVP		0.20	100	>5000 ^c	55	550
2		[Val ⁴]dDAVP		0.05	89	>20000 ^c	480	7000
3	Ph(4-OH)	CH ₂ OH	S	0.10	92	>10000 ^c	520	2300
4	Ph(4-OH)	CH ₂ OH	R	0.08	93	>12000 ^c	720	2000
5	Ph(4-Cl)	CH ₂ OH	R	0.10	102	>10000 ^c	1400	>100000 ^c
6	Ph(4-Cl)	CH ₂ OH	R	0.08	115	>12000 ^c	800	>120000 ^c
7	Ph(4-Cl)	CH ₂ OH	S	0.14	96	>7100 ^c	2700	>71000 ^c
8	Ph(4-Cl)	CH ₂ OH	R	0.35	102	>2800 ^c	620	>28000 ^c
9	Ph(4-Me)	CH ₂ OH	R	0.39	104	>25000 ^c	2000	>25000 ^c
10	Ph(4-Et)	CH ₂ OH	S	0.35	110	>2800 ^c	>28000 ^c	>28000 ^c
11	Ph(4-Cl)	H	NA	0.31	89	>3200 ^c	450	>32000 ^c
12	Ph(4-Cl)	H	NA	0.07	104	>140000 ^c	1500	>140000 ^c
13	Ph(4-Cl)	H	NA	0.12	106	>83000 ^c	1100	>83000 ^c
14	Ph(4-OH)	H	NA	0.19	94	>5200 ^c	>52000 ^c	3400 ^c
15	Ph(4-Cl)	C(=O)NHMe	R	0.27	106	>3700 ^c	1200	>37000 ^c
16	Ph(4-Cl)	C(=O)NHEt	R	0.29	92	>3400 ^c	580	>34000 ^c
17	Ph(4-Cl)	C(=O)NHPr	R	0.33	92	>3000 ^c	750	>30000 ^c
18	Ph(4-Cl)	C(=O)NHcPr	R	0.23	86	>4300 ^c	2000	>43000 ^c
19	Ph(4-Cl)	C(=O)NH _i Pr	R	0.45	81	>2200 ^c	1200	>22000 ^c
20	Ph(4-Cl)	C(=O)NHBu	R	0.26	98	>38000 ^c	730	>38000 ^c
21	Ph(4-Cl)	C(=O)NH _i Bu	R	0.22	100	>45000 ^c	950	>45000 ^c
22	Ph(4-Cl)	C(=O)NEt ₂	R	0.25	102	>40000 ^c	2100	>40000 ^c
23	Ph(4-Cl)	C(=O)NHBzl	R	0.19	100	>52000 ^c	780	>52000 ^c
24	Ph(4-Cl)	C(=O)NHEt	R	0.10	103	>100000 ^c	1600	>100000 ^c

^a XY=CH₂S for analogues 3, 5, 7-24 and SCH₂ for 4, 6; Ar²=2-thienyl for 3-7, 9, 10, 12-24 and 4-fluorophenyl for 8, 11; R²=C(=NH)NH₂ for all new compounds except for 14 where R²=H; ^b Ratio EC₅₀(receptor)/EC₅₀(hV₂R); ^c No agonism up to 1 μM or 10 μM, the highest concentration tested.

First, the C-terminal group R¹ (-C(=O)-Gly-NH₂) in compound 2 was replaced with the hydroxymethyl function and the analogues were also modified in position 2, 3 and at the disulfide

bridge (compounds **3-10**). The disulfide bridge modifications (X, Y) as well as the configuration of the ¹³C chiral carbon had very little impact on potency and selectivity profiles. Compounds in this series were potent hV₂R agonists with **9** and **10** (Ar¹ = 4-alkylphenyl) being about 2-fold less potent at the V₂R than **1**. Analogues **3-10** displayed improved selectivity versus both the hV_{1b}R (all partial agonists) and hOTR except for the Tyr² compounds **3** and **4** (Ar¹ = 4-hydroxyphenyl) that were less selective vs. hOTR than **2**. Peptide **8** where Ar² = 4-fluorophenyl was less potent as an hV₂R agonist than its 2-thienyl counterpart **5**.

Next, we investigated if the substituent R¹ is actually required to preserve high agonistic potency at the hV₂R (compounds **11-14**). Compound **12** (R¹ = H, Ar¹ = 4-chlorophenyl, Ar² = 2-thienyl) was found to be very potent and considerably more selective than **2**. The Thz⁷ modification (Z = S) was found to be neutral in this series as analogues **12** and **13** had very similar pharmacological profiles. In addition we demonstrated that the guanidino function (R² = -C(=NH)NH₂) does not appear to be essential to preserve hV₂R agonism (compound **14**, R² = H). Lastly, we explored if the C-terminal glycine amide could be replaced with alkyl groups (R¹ = -C(=O)-NR³R⁴; compounds **15-24**). Based on the initial results the cyclic part of the new analogues was fixed with the consensus structure (Ar¹ = 4-chlorophenyl, Ar² = 2-thienyl, X = CH₂, Y = S). Somewhat surprisingly a variety of R³, R⁴ substituents were well tolerated and rather shallow SAR was observed. The isopropyl compound **19** (R³ = iPr, R⁴ = H) was the least potent V₂R agonist in this subset. The Thz⁷ modification was advantageous as exemplified by a 3-fold increase in potency for peptide **24** (Z = S) vs. **16** (Z = CH₂). Double alkylation, e.g. compound **22** (R³ = R⁴ = Et), was also well tolerated while maintaining excellent selectivity vs. the related receptors.

In conclusion, a series of novel C-terminally truncated dDAVP analogues with improved *in vitro* pharmacological profile has been identified. The novel compounds retain the potent V₂ receptor agonism activity of dDAVP, **1**. Compounds **5-13** and **15-24** display substantially improved selectivities vs. hV_{1a}R, hV_{1b}R and hOTR as compared to **1** and **2**.

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